**Dear Editor,**

**I would like to thank you for your insightful comments and concerns. Detailed below are the changes made in response to each comment individually. Your comments will be copied verbatim in black, and our response will be in blue**

So invitro, invivo and the P.aeruginosa pneumonia methods are different? Needs more clarity on this part.

Edited as suggested.

What is the significance of this with respect to phagocytosis?

We believe, explaining an additional method of bacterial clearance will have more advantage.

Please revise the Introduction to include all of the following:

Edited as suggested.

Citation.

Relevant citations are added.

Need to bring out the clarity in the protocol with respect to in vivo and in vitro

Relevant editions are made.

Age, sex, strain of mice used?

Required information is added.

How small of an incision? What was used for the incision?

Required information is added.

So everytime you are withdrawing and re-infusing it back. Or you are just infusing 3ml of PBS. How do you maintain the sterility in this case?

In our hand, this method gives higher cell number. Every time we withdraw and re-infuse back 1 mL PBS, and transfer the collected BALF to a sterile tube.

Please mention that the PBS above is called BALF here. Volume collected?

Required details are added.

How do you call it as alveolar macrophages at this stage? How about cell debris.

Required changes are made

Do you wash before culturing? Do you filter?

Required details are added.

What kindo of beads are these.

Bead details are already mentioned.

Do you check for any macrophage-based marker at this stage? How do you get rid of non-specific bindings.

In our experience, under resting conditions, more than 95 % of cells are macrophages and they look very different from other cells. We have also tried using cell sorting by flow cytometry. But, under resting condition, differential staining and microscopy is good enough to identify AMs.

Citation

New references are included.

Wouldn’t this lyse the cells with RBC as well? Why can’t just use PBS instead? Volume of the buffer used?

Care should be taken not to exceed washing time more than 1 minute. This will only wash outside RBCs. same is discussed in discussion and relevant reference is added.

What are the culture conditions? How many cells per dish?

Required details are added.

Grow how? Also do you grow on agar or broth?

Required details are added.

How is this done?

Required details are added.

Conditions speed temp time etc ?

Required details are added.

You cannot call this Model since more specific studies are needed. Please reword.

Corrections are made.

What is the desired CFU and how do you calculate this. Please include a one liner note for this.

A note is added.

This is on a different set of mice?

Yes.

Please include a one-liner title for the figures representing all panels together.

A one-liner title is added.

As we are a methods journal, please revise the Discussion to explicitly cover the following in detail in 3-6 paragraphs with citations:

Changes are made.

The editor has formatted the manuscript to match the journal's style. Please retain the same.

Please address all the specific comments marked in the manuscript.

Specific comments are addressed.   
3. Once done please ensure that the protocol is no more than 10 pages and the highlights are no more than 2.75 pages including heading and spacings.  
4. Figure 3A would benefit from borders on the microscopic images. It is difficult to separate the different images visually.

Changes are made.

5. Please provide copyright permissions for the Figures if they are to be re-used in publication.

The copy of copyright permissions we submitted before covers both text and figures to be reused.